

ISOLATION AND CHARACTERIZATION OF URSANE AND OLEANANE TYPE TRITERPENES FROM THE ROOTS OF *STREPTOCAULON TOMENTOSUM*. WIGHT & ARNOTT (Utrifp*le) (ASCLEPIADACEAE) IN MYANMAR

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Abstract

The aim of this paper is to study the NMR spectroscopic characterization of seven triterpenes from *Streptocaulon tomentosum* Wight & Arnott (Asclepiadaceae). Four ursane type triterpenes and three oleanane type triterpenes were isolated from the roots of *Streptocaulon tomentosum*. by column chromatography and identified by NMR spectroscopy. Comparative studies on NMR spectra of ursane type and oleanane type triterpenes were done. Four ursane type triterpenes were identified as α -amyrin acetate (compound 2), $2\alpha,3\alpha,23$ -trihydroxy-urs-12-en-28-oic-acid (compound 3), $2\alpha,3\beta$ -dihydroxy-urs-12-en-28-oic-acid (compound 4), and $2\alpha,3\beta,23$ -trihydroxy-urs-12-en-28-oic-acid (compound 6). Three oleanane type triterpenes were identified as β -amyrin acetate (compound 1), $2\alpha,3\beta$ -dihydroxy-olean-12-en-28-oic-acid (compound 5) and $2\alpha,3\beta,23$ -trihydroxy-olean-12-en-28-oic-acid (compound 7).

Keywords: *Streptocaulon tomentosum*, Asclepiadaceae, ursane, oleanane, triterpenes

Introduction

The roots of *Streptocaulon tomentosum* are used in Myanmar in traditional medicine for the treatment of anticancer, dysentery and stomach-ache, and the leaves are used externally for the treatment of snake poisoning and abscesses. The triterpenoids, having a C₃₀ skeleton, constitute a large, diverse group of natural products derived from squalene or, in the case of 3β -hydroxy triterpenoids, the 3S-isomer of squalene 2,3-epoxide. In excess of 4000 triterpenoids have been isolated so far and more than 40 skeletal types have been identified. Oleananes and ursanes often occur together and, in the past decade, have been reported from a wide range of families including the Araliaceae, Asclepiadaceae, Bignoniaceae, Cactaceae, Campanulaceae, Celastraceae, Compositae, Ericaceae, Fagaceae, Labiateae, Leguminosae, Phytolaccaceae, Primulaceae, Rosaceae, Rubiaceae, Sapotaceae, Theaceae, Umbelliferae and Urticaceae (Dey & Harborne, 1991). The biological activities of triterpenoids and triterpenoid saponins are immunostimulation (Press *et al.*, 2000), anti-tumor-promoting activity (Konoshima & Takasaki, 2000), anti-inflammatory activity and anti-insect activity (Connolly & Hill, 2002). This paper focused on the NMR spectroscopic characterization of ursane and oleanane types triterpenes isolated from the roots of *Streptocaulon tomentosum* Wight and Arn. (Asclepiadaceae).

Materials and Methods

Sample Collection and Identification of Plant Samples

Streptocaulon tomentosum Wight and Arn. (Asclepiadaceae), roots collected in May 2002 from Mawlamyine Township, Myanmar by Dr Daw Hla Ngwe. The species was identified by Prof. Dr Aung Aung Min, Department of Botany, University of Yangon. A voucher specimen of the clamberer (No. Y.H.V. 1004) is deposited in University of Yangon.

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Spectral Studies

1D NMR spectra (^1H , ^{13}C) were recorded on a Varian Mercury 300 at 300.94 MHz for ^1H , and at 100.57 MHz for ^{13}C NMR. Chemical shifts in ppm were referenced to internal TMS ($\delta = 0$) for ^1H and $\text{C}_5\text{D}_5\text{N}$ (δ 149.81, 135.48, 123.50 ppm) for ^{13}C , respectively.

Isolation of Triterpenes from the Roots of *Streptocaulon tomentosum*

Dried powdered root of *Streptocaulon tomentosum* (Asclepiadaceae) (1 kg) was extracted with 80% EtOH (1 L \times 3) for one week. The solvent was evaporated to the remaining water layer. Then the water layer was successively partitioned between organic solvents (*n*-hexane, ethyl acetate, *n*-butanol 300 mL \times 3) and water. The *n*-hexane fraction (48 g) was chromatographed over silica gel 60 (70-230 mesh, Merck), using a stepwise gradient of *n*-hexane: ethyl acetate (9.5:0.5, 9:1,2:1, increasing polarity) to give four fractions. Fraction 1 (8 g) and fraction 2 (4 g) were rechromatographed on silica gel 60 column (230-400 mesh, column size id 2 cm \times 60 cm) with the solvent system *n*-hexane: chloroform (3:1) and (9:1-4:1) to give β -amyirin acetate (**1**), and α -amyirin acetate (**2**). The ethyl acetate fraction (12 g) was separated on a silicagel column (70-230 mesh, column size id 3 cm \times 60 cm) and eluted with *n*-hexane: ethyl acetate: methanol (increasing polarity 9:1:0, 7:3:0.5,to pure MeOH) to give 23 fractions (each about 300 mg). Fractions 5-11 were rechromatographed on silica gel 60 (230-400 mesh, column size 1.5 cm \times 40 cm) using CHCl_3 : MeOH (9.5:0.5; 4.7:0.3) to give compounds **3-7**.

Identification of Ursane and Oleanane Types Triterpenes

The structures elucidations were determined by ^1H NMR, ^{13}C NMR, ESI-MS, FT-IR, GC-MS and confirmed by comparison with the literature values (Matsunaga *et al.*, 1988; Chen *et al.*, 1993; Hisham *et al.*, 1993; De Pascual Teresa *et al.*, 1987).

Results and Discussion

Structure Elucidation of Ursane and Oleanane Type Triterpenes

The *n*-hexane fraction was repeatedly chromatographed on a silica gel column and β -amyirin acetate (**1**), and α -amyirin acetate (**2**). The structures elucidations were determined by ^1H NMR, ^{13}C NMR, ESI-MS, GC-MS and confirmed by comparison with the literature values.

Compound 1: β -amyirin acetate

colourless needles Yield: 100 mg, 0.01%

mp.: 242-243 °C

$[\alpha]_D$: + 80.1 ° (c = 1.10, CHCl_3)

TLC: R_f = 0.47 (system T₁, violet colour with vanillin/ H_2SO_4 , inactive under UV)

IR: (KBr), ν_{max} = 1730, 1635, 1240, 812 cm^{-1}

^1H NMR: (300 MHz, CDCl_3): δ 0.84 (3H, *s*, H-28), 0.88 (12H, *s*, H-23, 24, 29, 30), 0.98 (6H, *s*, H-25, 26), 1.14 (3H, *s*, H-27), 2.07 (3H, *s*, OAc), 4.54 (1H, *dd*, *J* 11.6 Hz, H-3 α), 5.21 (1H, *t*, *J* 3.5 Hz, H-12)

GC-MS: RT = 18.03 min, 468 $[\text{M}]^+$, 453 (33), 408 $[\text{M}-\text{HOAc}]^+$ (5), 393 (40), 281 (70), 218 (100), 203 (68), 69 (90)

EI-MS: (70 ev) *m/z* (rel. int): 468 $[\text{M}]^+$, 453 (2), 408 $[\text{M}-\text{HOAc}]^+$ (5), 218 (100), 203 (20)

Compound 2: α - amyrin acetate

Colourless needles, Yield: 1.5 g, 0.15%

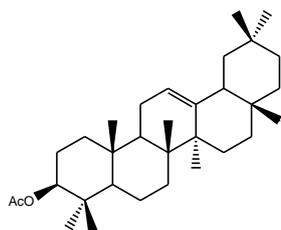
mp.: 243 °C

[α]_D: + 76 ° (c = 1.0, CHCl₃)TLC: R_f = 0.47 (system T₁, violet colour with vanillin/H₂SO₄, inactive under UV)IR: (KBr), ν_{\max} = 1730, 1380, 1370, 1250, 1030, 1000, 985, 960 cm⁻¹

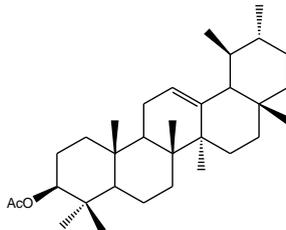
¹H NMR: (300 MHz, CDCl₃): δ 0.79 (3H, s, H-28), 0.88 (12H, s, H-23, 24, 29, 30), 0.98 (3H, s, H-26), 1.01 (3H, s, H-25), 1.07 (3H, s, H-27), 2.05 (3H, s, OAc), 4.50 (1H, dd, *J* 9.7 Hz, H-3 α), 5.12 (1H, t, *J* 3.6 Hz, H-12)

GC-MS: RT = 18.85 min, 468 [M]⁺, 453 (45), 408 [M- HOAc]⁺ (38), 393 (40), 281 (100), 218 (80), 203 (30), 69 (98)

EI-MS: (70 ev) *m/z* (rel. int): 468 [M]⁺, 453 (10), 408 [M- HOAc]⁺ (20), 218 (100), 203 (10)



Compound 1
(Oleanane type)



Compound 2
(Ursane type)

Compound **3-7** were isolated from fraction 5-11 of the ethyl acetate extract after repeated column chromatography on silica gel. Identification of these known compounds was based on 1D and 2D NMR, MS and comparison of their spectroscopic data with literature values (Sashida *et al.*, 1992; Kojima & Ogura, 1986; Yaguchi, 1988).

In the compounds **3, 6** and **7** the molecular formula is C₃₀H₄₈O₅ by means of HR-ESI-MS. In compound **4** and **5** the molecular formula is C₃₀H₄₈O₄. Their mass spectra present ions at *m/z* 248 resulting from the retro-Diels-Alder fragmentation characteristic of the ursane and oleanane skeletons. Furthermore they possess an ion at *m/z* 203 characteristic of Δ^{12} -triterpenoids (Budzikiewicz *et al.*, 1963). In the ¹H NMR (C₅D₅N) spectra of these compounds, the signal of H-18 permitted the distinction between the oleanane and ursane skeletons. The H-18 signal appears at δ 2.6 ppm in the ursane skeleton and at δ 3.3 ppm in the oleanane skeleton. The proton signals of H-29 and H-30 in the ursane skeletons appears as a doublet, but in oleanane as a singlet. The chemical shifts of C-12 and C-13 (δ 125 and 139 ppm in ursane, δ 122 and 144 ppm in oleanane) and H-12 (δ 5.20-5.4 ppm) suggests that these compounds are Δ^{12} -unsaturated triterpenoids. The ¹³C NMR spectra (table. 1) clearly exhibited the difference in the chemical shifts of C-12, C-13, C-17, C-18, C-19, C-20, C-22, C-27, C-29 and C-30 between the ursane group (**3, 4, 6**) and the oleanane group (**5, 7**). The coupling constant of H-3 (*J* 2.3 Hz) in **3** suggested that two OH groups at C-2 and 3 were at the *cis* position. Besides, the ROESY correlation between H-3 and H-2, H-23, H-24 also confirmed the β -configuration of H-2 and H-3. However, the coupling constant of H-3 (*J* 9.4 Hz) in **4-7** showed that the two OH groups at C-2 and C-3 were in *trans* position and there was no ROESY correlation between H-3 and H-2 (see **HMBC, ROESY** in table 2).

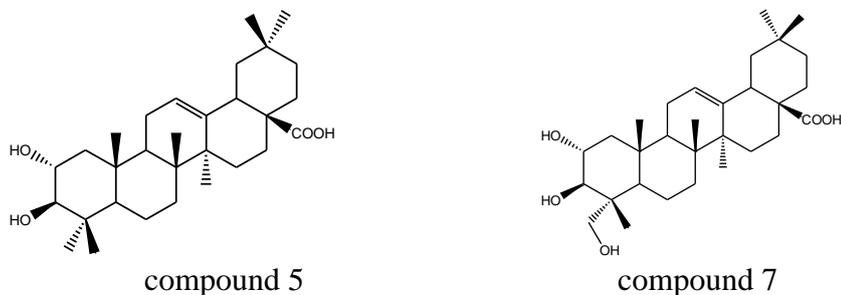


Figure 1 Oleanane type triterpenes

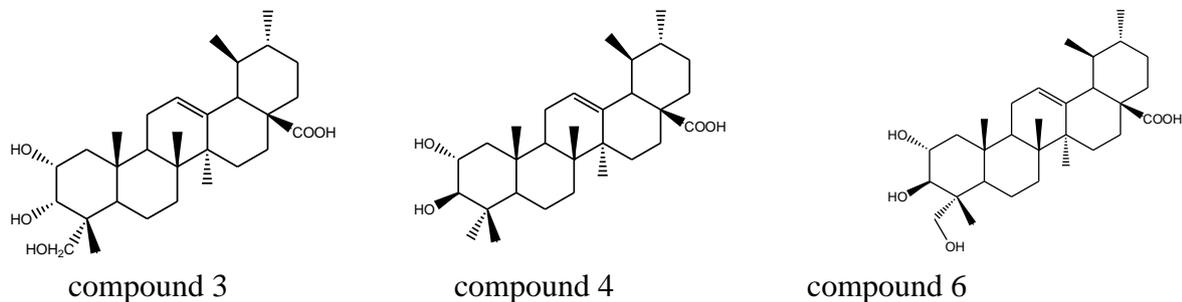


Figure 2 Ursane type triterpenes

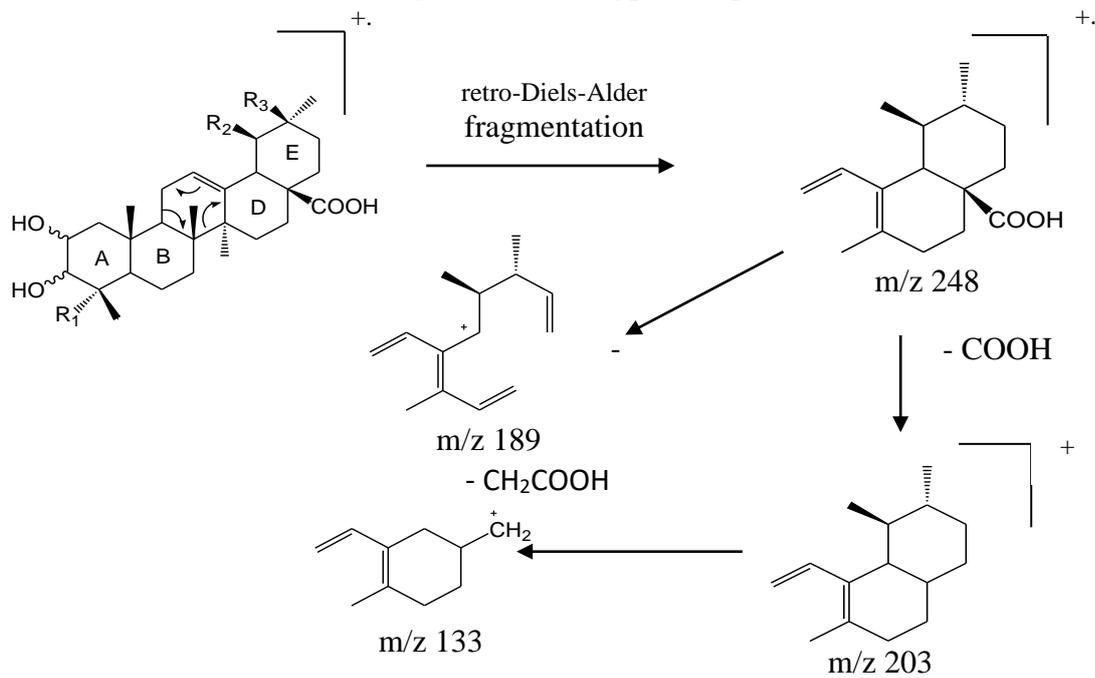


Figure 3 Proposed EI mass spectral fragmentation of triterpenoids 1-7

Table 1 ^{13}C NMR spectral data of triterpenoid 2-7
(300, 500 MHz, 2 in CDCl_3 ; 3-5 in $\text{C}_5\text{D}_5\text{N}$; 6-7 in CD_3OD)

C-Atom	δ_{C} [ppm]					
	2	3	4	5	6	7
1	38.5	42.8	47.9	47.7	48.0	47.9
2	23.4	66.2	68.6	68.6	69.7	69.7
3	80.9	78.9	83.8	83.8	78.1	78.1
4	37.7	41.9	40.0	40.0	44.1	44.1
5	55.3	43.5	55.9	55.9	48.1	48.1
6	18.3	18.3	18.8	18.8	19.1	19.1
7	32.9	33.2	33.2	33.2	33.6	33.6
8	39.7	40.1	39.8	39.8	40.8	40.5
9	47.7	47.9	48.1	48.1	48.2	48.2
10	36.8	38.3	38.4	38.5	39.0	39.0
11	22.8	23.7	23.7	23.7	24.5	24.5
12	124.2	125.5	125.5	122.4	126.6	123.4
13	139.5	139.3	139.3	144.7	139.8	145.4
14	42.1	42.5	42.5	42.2	43.4	43.0
15	28.2	28.6	28.6	28.2	29.1	28.8
16	26.7	24.9	24.9	23.9	24.6	24.0
17	33.8	48.0	48.0	46.6	48.9	47.6
18	59.0	53.5	53.5	41.9	54.3	42.7
19	39.7	39.4	39.4	46.4	40.4	47.2
20	39.7	39.4	39.3	30.9	40.4	31.8
21	31.3	31.0	31.0	33.5	31.6	33.3
22	41.6	37.4	37.4	34.2	38.1	34.9
23	28.1	71.2	29.4	29.3	66.2	66.2
24	15.8	17.8	17.7	17.7	13.9	13.9
25	14.2	17.1	16.9	16.9	17.5	17.5
26	16.8	17.5	17.5	17.7	17.8	17.7
27	17.6	23.8	23.9	26.1	24.1	26.5
28	28.8	179.9	179.9	179.9	181.7	181.5
29	23.3	17.5	17.5	33.2	17.7	33.6
30	21.4	21.3	21.4	23.7	21.6	24.0
COMe	21.5					
CO	170.8					

Table 2 NMR data of triterpenoids 3-7 (500 MHz, 3-5 in C₅D₅N; 6-7 in CD₃OD)

H-Atom	δ_H [ppm]					HMBC	ROESY
	3	4	5	6	7	(3-7)	(3-7)
1	1.82, 1.94 (<i>m</i>)	1.28, 2.26 (<i>m</i>)	1.28, 2.26 (<i>m</i>)	0.88, 1.96 (<i>m</i>)	0.88, 1.96 (<i>m</i>)	C-2, 3, 25	
2	4.289 (<i>m</i>)	4.115 (<i>ddd</i> , 11/9.4/4.4)	4.115 (<i>ddd</i> , 11/9.4/4.4)	3.687 (<i>m</i>)	3.687 (<i>m</i>)	C-3	H-3 (in 3), 25, 24
3	4.168 (<i>d</i> , 2.3)	3.420 (<i>d</i> , 9.4)	3.420 (<i>d</i> , 9.4)	3.350 (<i>dd</i> , 9.7/2.4)	3.350 (<i>dd</i> , 9.7/2.4)	C-24, 4, 1	H-2 (in 3), 23, 24
5	2.02- 2.08 (<i>m</i>)	1.04 (<i>m</i>)	1.04 (<i>m</i>)	1.28 (<i>m</i>)	1.28 (<i>m</i>)	C-25, 10, 4	
6	1.34, 1.60 (<i>m</i>)	1.36, 1.54 (<i>m</i>)	1.36, 1.54 (<i>m</i>)	1.38 (<i>m</i>)	1.38 (<i>m</i>)	C-25, 5	
7	1.34, 1.72 (<i>m</i>)	1.84, 2.04 (<i>m</i>)	1.84, 2.04 (<i>m</i>)	1.54, 1.74 (<i>m</i>)	1.54, 1.74 (<i>m</i>)	C-26, 8, 14	
9	1.94 (<i>m</i>)	1.76 (<i>m</i>)	1.76 (<i>m</i>)	1.66 (<i>m</i>)	1.66 (<i>m</i>)	C-25, 11, 10, 8, 5,	H-25, 26
11	1.96- 2.08 (<i>m</i>)	1.98 (<i>m</i>)	1.98 (<i>m</i>)	1.94 (<i>m</i>)	1.94 (<i>m</i>)	C-12, 13, 9, 8, 10	
12	5.480 (<i>brs</i>)	5.476 (<i>m</i>)	5.476 (<i>m</i>)	5.242 (<i>m</i>)	5.242 (<i>m</i>)	C-11, 14, 9, 18, 13	H-18, 29
15	1.14- 2.36 (<i>m</i>)	1.18, 2.36 (<i>m</i>)	1.18, 2.36 (<i>m</i>)	1.08 (<i>m</i>)	1.08 (<i>m</i>)	C-27, 26, 16, 8, 14	
16	1.98- 2.06 (<i>m</i>)	2.00, 2.12 (<i>m</i>)	2.00, 2.12 (<i>m</i>)	1.94 (<i>m</i>)	1.94 (<i>m</i>)	C-15, 17	
18	2.626 (<i>br d</i> , 11.3)	2.641 (<i>br d</i> , 11.4)	3.314 (<i>dd</i> , 13.9, 4.0)	2.202 (<i>d</i> , 11.2)	2.849 (<i>dd</i> , 13.6, 3.9)	C-9, 16, 19, 20, 14, 17, 12, 13, 28	H-29, 20 (in ursane type), 12
19	1.42 (<i>m</i>)	1.46 (<i>m</i>)	1.28, 1.80 (<i>m</i>)	1.38 (<i>m</i>)	1.14, 1.70 (<i>m</i>)	C-29, 30, 22, 20, 17, 18	
20	1.00 (<i>m</i>)	1.04 (<i>m</i>)		0.98 (<i>m</i>)		C-30, 21 (in ursane type)	
21	1.34, 1.44 (<i>m</i>)	1.40 (<i>m</i>)	1.36, 1.56 (<i>m</i>)	1.36, 1.52 (<i>m</i>)	1.28, 1.66 (<i>m</i>)	C-29, 30, 22, 20, 17,	
22	1.96 (<i>m</i>)	1.98 (<i>m</i>)	1.18, 1.46 (<i>m</i>)	1.66 (<i>m</i>)	1.20, 1.40 (<i>m</i>)	C-17, 28	
23	3.77, 3.94 (<i>d</i> , 10.8)	1.291 (<i>s</i>)	1.291 (<i>s</i>)	3.261 (<i>d</i> , 11.0)	3.261 (<i>d</i> , 11.0)	C-24, 2, 3, 4, 5	H-3
24	0.87 (<i>s</i>)	1.092 (<i>s</i>)	1.092 (<i>s</i>)	0.692 (<i>s</i>)	0.690 (<i>s</i>)	C-25, 5, 23, 3	4, H-25,2, 3, 23
25	1.00 (<i>s</i>)	0.991 (<i>s</i>)	0.991 (<i>s</i>)	1.042 (<i>s</i>)	1.028 (<i>s</i>)	C-24, 11, 10, 4, 5, 9, 1	H-24, 26, 3
26	1.07 (<i>s</i>)	1.060 (<i>s</i>)	1.032 (<i>s</i>)	0.846 (<i>s</i>)	0.813 (<i>s</i>)	C-25, 7, 8, 14, 9	
27	1.14 (<i>s</i>)	1.220 (<i>s</i>)	1.275 (<i>s</i>)	1.132 (<i>s</i>)	1.175 (<i>s</i>)	C-15, 8, 14, 18, 12, 13	
29	0.965 (<i>d</i> , 6.4)	0.995 (<i>d</i> , 4.9)	0.954 (<i>s</i>)	0.865 (<i>d</i> , 6.4)	0.907 (<i>s</i>)	C-19, 20	
30	0.925 (<i>d</i> , 6.2)	0.960 (<i>d</i> , 5.9)	1.014 (<i>s</i>)	0.967 (<i>d</i> , 6.0)	0.941 (<i>s</i>)	C-21, 19, 20	

Conclusion

Oleanane and ursane type triterpenes are rich in the roots of *Streptocaulon tomentosum* Wight. Literature survey showed that oleanane and ursane tetraterpenes possess anti-tumor-promoting activity, anti-inflammatory activity and anti-insect activity. So the roots of *Streptocaulon tomentosum* Wight may be useful for the treatment of anticancer.

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References

- Budzikiewicz, H., Wilson, J. M., Djerassi, C. (1963). "Mass Spectrometry in Structural and Stereochemical Problems. XXXII.1 Pentacyclic Triterpenes", *J. Am. Chem. Soc.* vol. 85, pp. 3688.
- Connolly, J. D., Hill, R. A. (2002). "Triterpenoids", *Nat. Prod. Rep.* vol. 19, pp. 494-513.
- De Pascual Teresa, J., Urones, J. G., Marcos, I. S., Basabe, P., Sexmero Cuadrado, M. J., Fernandez Moro, R. (1987). "Triterpenes from *Euphorbia broteri*", *Phytochemistry*. vol. 26, pp. 1767-1776.
- Dey, P. M., Harborne, J. B. (1991). *Methods in Plant Biochemistry*. vol. 7, pp. 341-342.
- Hisham, A., Kumar, G. J., Fujimoto, Y., Hara, N. (1995). "Salacianone and salacianol: two triterpenes from *Salacia beddomei*", *Phytochemistry*. vol. 40, pp. 1227.
- Kojima, H., Ogura, H. (1986). "Triterpenoids from *Prunella vulgaris*", *Phytochemistry*. vol. 25, pp. 729-733.
- Konoshima, T., Takasaki, M. (2000). "Cancer-chemopreventive effects of natural sweeteners and related compounds", *Stud. Nat. Prod. Chem.* vol. 24, pp. 607.
- Matsunaga, S., Tanaka, R., Akagi, M. (1988). "Triterpenoids from *Euphorbia maculata*", *Phytochemistry*. vol. 27, pp. 535-537.
- Press, J. B., Reynolds, R. C., May, R. D., Marciani, D. J. (2000). "Structure/function relationships of immunostimulating saponins". *Stud. Nat. Prod. Chem.* Vol. 24, pp. 131-174.
- Sashida, Y., Ogawa, K., Mori, N., Yamanouchi, T. (1992). "Triterpenoids from the fruit galls of *Actinidia polygama*" *Phytochemistry* vol. 31, pp. 2801-2804.
- Yaguchi, Y., Sakurai, N., Nagai, M., Inoue, T. (1988). "Constituents of *Myrica rubra*. III: Structures of Two Glycosides of Myricanol", *Chem. Pharm. Bull.* vol. 36, pp. 1419-1424.